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Assessment of Soil Parameters and Microbes in Amaravathi River Bed Area, Karur District, Tamil Nadu, India

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Abstract

In the present study, the physico-chemical properties of soil samples of agricultural region collected from Amaravathi River basin were analyzed. Parameters like soil pH, turbidity, total dissolved solids, total hardness, total alkalinity, calcium, magnesium, nitrate, sulphate, chloride, fluoride, phosphate, iron, manganese, biological oxygen demand and chemical oxygen demand for the soil samples were carried out. Different values were observed in the different physico-chemical parameters due to the soil quality in different sampling locations. Values of pH, Chloride, Fluoride, Sulphate, BOD and COD concentration in the soil samples also show wide variations. Microbiological characteristics of soil samples were carried out among which, bacillus and pseudomonas species were the predominant bacteria genera isolated.

Keywords: Amaravathi River; Karur; Microbes; Physico-chemical properties.

1. INTRODUCTION

Soil is a dynamic medium created of minerals, organic matter, water, air and living organisms like bacteria and earth worms, and its continuous properties charging due to farming factors such as parent material, time, climate and the organisms (Nwachokorm *et al.* 2009). Healthy soil consists of approximately 40 % mineral, 23 % water, 23 % air, 6 % organic material and 8% living organisms. A study of soil profile supplemented by physical, chemical and biological properties of the soil will give full picture of soil fertility and productivity. Good productive soil builds the foundation for any successful cropland (Galal M. Zaiad, 2001). The worldwide significant decline in soil productive capacity through physical, chemical,

biological properties and contamination is by inorganic and organic chemicals (Abu Ahmed Mokammel Haque, 2007). Soil as a module of the terrestrial ecosystem fulfills many activities that are important for plant growth. The qualities that are used to assess the ability of soil to fulfill storage of plant available water, supply of possible oxygen to roots, storage of nutrients essential to plant growth, provision of favorable conditions of seeding establishment and these functions include pH, cation exchange capacity and soil depth (Nwachokorm *et al.* 2009).

The most unadorned and widespread difficulties facing the agriculture department is the degradation of soil quality due to salinity. Almost 40% of the land surface in world is pretentious by salinity related problems (Pandeeswari and Kalaiarasu, 2012). India is a leading country using large number of fertilizers instead of manures to this crop productivity increases

*A. Jafar Ahamed Tel.: +91-9003576896 E-mail: agjafar@yahoo.co.in. but the quality of soil decreases. Pollution in soil and water is strictly related to human activities such as industry, burning fossil fuels, mining and metallurgical processes and their waste disposals (Ladwani Kiran, 2012). It is real time to carry out the physico-chemical analysis of soil and river sediments.

Micro organisms in the soil contribute to the improvement and maintenance of soil quality. The dynamic nature of soil organisms makes them sensitive to nature or management – related soil changes and thus makes excellent assessors of soil quality. Soil microbes perform some beneficial functions as well as some deterimental impacts to environment. In the study area, adverse effects on soil health and soil quality arise from nutrient imbalance in soil, excessive fertilization, soil pollution from industrial units and soil loss processes. The aim of the study therefore was to provide the baseline data of soils for its management for sustainable agriculture.

2. GEOLOGY OF THE STUDY AREA

Amaravathi River is a tributary of Cauvery River in Coimbatore and Tiruppur, Tamil Nadu, South India. The 175 km long Amaravathi River begins at the Kerala/Tamil Nadu border at the bottom of Manjampatti Valley between the Annamalai Hills and the Palani hills in Indira Gandhi Wildlife Sanctuary and National Park in Tiruppur. It descends in a northerly direction through Amaravathi Reservoir and Amaravathi Dam at Amaravathinagar. It is joined by the Kallapuram River at the mouth of the Ajanda valley in Udumalaipettai, and it merges with the river Cauvery at Thirumukkudal, about 10 km from Karur. Amaravathi River irrigates over 60,000 acres of agricultural lands in Coimbatore, Erode and Karur districts and fulfills the drinking water requirements of these regions. The Amaravathi River and its basin, especially in the vicinity of Karur, are heavily used for industrial processing water and waste disposal and as a result are severely polluted due to large amount of textile dyeing and bleaching units.

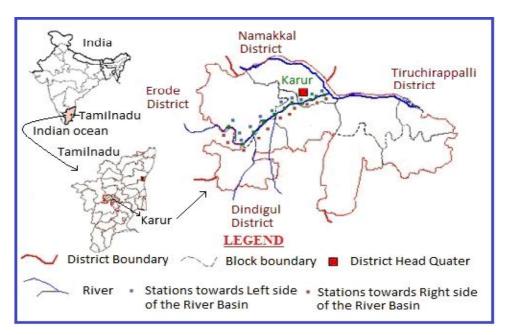


Fig. 1: Map of the study area showing sampling sites

Table 1. Physico-chemical Characteristics of the Soil Samples

COD	274	251	245	269	236	283	249	261	273	260	342	212	275	260	245	272	256	230	259	235	250	240	264	270	196	245	278	256	223	296	267	215	183	249	160	274
BOD	340	290	365	285	316	386	341	374	322	300	289	347	360	245	265	270	250	240	260	254	271	280	244	236	342	298	316	350	379	341	326	384	265	421	356	372
Мп	0.05	0.03	0.06	0.1	0.08	0.03	0.07	90.0	0.05	0.05	0.00	90.0	0.3	0.07	0.1	0.09	0.05	90.0	0.08	0.03	0.1	0.03	0.05	0.07	0.02	0.08	0.07	0.05	0.07	0.02	0.05	0.09	0.04	0.03	0.01	0.03
F e	80.0	0.05	0.1	0.15	0.1	90.0	0.3	0.1	0.5	0.7	0.7	0.1	0.4	0.2	0.1	90.0	0.00	0.02	0.3	0.7	80.0	0.3	0.1	0.7	0.3	0.1	0.00	0.3	80.0	0.4	0.1	9.0	0.5	0.7	1.0	0.7
N 0 3	46	49	52	58	49	47	46	51	45	52	54	46	32	4 8	55	36	54	45	42	37	52	45	44	50	63	45	74	39	19	48	85	69	41	78	49	52
P 0 4	90.0	0.1	0.04	0.07	0.01	0.02	90.0	80.0	0.02	0.01	0.05	0.01	0.03	0.01	0.02	0.02	0.01	0.03	0.05	0.02	0.03	0.01	0.01	0.02	0.09	0.03	0.01	0.04	0.02	0.01	0.07	0.02	90.0	0.01	0.03	0.07
SO4	146	126	203	158	194	186	175	137	215	120	164	193	155	167	142	150	145	160	162	148	150	140	143	163	150	137	162	141	153	140	210	184	208	195	178	185
H	1.0	1.1	1.5	1.3	1.1	0.09	1.2	0.05	9.1	1.5	1.4	1.5	1.5	1.9	1.0	1.3	1.2	1.6	0.7	1.0	6.0	1.0	1.4	8.0	2.0	3.2	1.3	2.6	1.0	1.8	1.2	1.7	1.4	2.1	1.1	2.4
CI	245	268	236	265	250	276	296	257	271	260	266	258	280	230	245	260	235	270	240	245	260	272	250	225	259	163	296	278	249	251	271	237	310	284	244	186
Mg																																39	73	82	51	70
C a																									96							82	41	24	39	57
TA	156	196	245	268	189	178	132	275	261	150	253	200	210	182	200	215	210	190	200	195	180	190	192	105	230	180	215	230	190	255	196	237	247	154	249	168
ΗI	236	275	253	274	195	214	256	217	248	210	185	274	200	190	215	222	205	212	215	230	180	200	200	205	250	192	236	257	165	199	186	240	257	186	146	214
TDS	563	500	516	465	584	624	575	319	652	460	535	584	540	450	500	490	460	510	530	495	515	500	520	525	009	450	560	610	570	550	580	540	200	565	480	504
EC	186	831	945	871	1020	1198	964	702	1247	831	887	1062	774	719	863	876	793	206	920	887	902	975	918	964	1112	853	1005	1264	1037	930	897	955	923	1010	863	816
Ηď	7.40	8.02	7.85	7.20	8.12	7.53	8.00	7.50	7.34	7.78	8.27	8.54	7.25	7.95	8.23	7.13	7.67	7.97	7.45	8.11	8.39	8.26	7.78	8.35	7.38	7.54	7.72	7.43	7.25	7.21	7.82	7.65	8.34	8.12	7.88	7.98
Turbidity																									3.4									3.5	2.6	2.8
Sample Id	ST 1	SL 2	ST 3	SL 4	ST S	9 TS	L TS	8 TS	6 TS		SL 11		-	_	_		1			C	C	C	C	CI	C	C	~	~	C	•	~		~	3	RS 35	CC)

All the values are expressed in mg/L except pH and EC in iS/cm.

Amaravathi watershed (Fig. 1) in Karur district lies between 10° 77' and 10° 95 N' and 77° 92' and 78° 23 E'. The lithology of the study area is characterized with older Pre-Cambrian basement and younger alluvia. Rocks of the khondalite group including garnet-sillimanite schists and gneisses, calc-granulites with crystalline schists and gneisses, calc-granulites with crystalline limestone pockets and quartzite are well exposed. Quartz of very fine quality is found in many places, especially in the village of Vaduvanampalli near Kodiyur and Kothapalayam near Aravakurichi. A major portion (90 %) of the soil of the Amaravathi basin is red sand. Karur has average annual rainfall of 25.04 in inches and average number of rainy days in a year is 40.

3. MATERIALS & METHOD

3.1 Soil Sampling

Soil samples were collected from thirty six different places (including river bed soil) of Amaravathi river basin area of Karur district. Soil samples were collected from a depth of 5-10 cm during January 2013 into labeled sterile polythene bags and taken in ice-packed coolers to the laboratory for physico-chemical and microbiological analysis. For chemical measurements, the soil samples were air-dried and then sieved through a 2 mm sieve.

3.2 Physico-Chemical Analysis of Soil Samples

Aqueous extract of the samples were prepared by mixing 80 g of the air dried sample with 400 ml double distilled water in 500 ml beaker. Using hot plate magnetic stirrer the mixture was stirred for 30 minutes at 40 °C, and then the mixture was allowed to settle for one hour. Filtrates of soil-water slurry (1:5 w/v) were used for chemical analysis. The same procedure was adopted for each sample. Soil pH and electrical conductivity (EC) and total dissolved solids (TDS) were determined by using combined water quality multi-parameter probe Elico PE 138 (Jackson, 1973). The sum of the calcium (Ca) and magnesium (Mg) contents in the extract was determined by the Eriochrome Black-T titration method

and the Ca content was then subtracted from the sum to determine the Mg content. Total alkalinity (TA) was determined by titrating with 0.1 N HCl using methyl orange as indicator. Chloride (Cl) content of the soil samples was determined by Mohr's method (Washington, 1954). Turbidity and sulphate (SO₄²) were estimated using Nepholometer. Fluoride (F) was estimated colorimetrically by **SPADNS** [2-(p-sulphophenylazo)1, 8-dihydroxynapthalene-3, 6disulphonic acid trisodium salt), C₁₆H₀N₂O₁₁S₃Na₃]. Nitrate (NO₂) was analyzed by spectrophotometric determination at 520 nm. A value for biochemical oxygen demand (BOD) was obtained using the Winkler's titration method. Chemical Oxygen Demand (COD) was determined by dichromate open reflex method. Iron (Fe) and manganese (Mn) were analyzed using the Atomic Absorption Spectroscopy (AAS).

3.3 Microbiological Analysis of the Soil Samples

The soil microbiological analysis of the samples were carried out according to the methods of Rabah (2008) and Oyeleke and Manga (2008). The bacterial isolates were identified and characterized using standard biochemical tests (Cheesebrough, 2006). The tests employed include gram stain, motility, catalase, oxidase, methyl red, nitrate, Voges-Proskaeur, indole production, urease activity and citrate utilization tests.

4. RESULTS & DISCUSSION

4.1 Soil Physico-chemical Properties

The physico-chemical properties of the soil samples analyzed are presented in table 1. The turbidity was measured in the NTU units and the turbidity of the soil samples were ranged from 1.4 to 3.6 NTU. The pH in soil is the measure of hydronium ion activity in the soil solution. Soil pH influences many facts of crop production and soil chemistry, as well as availabilities of nutrients and toxic substances, activities and nature of microbial populations, and activities of certain pesticides (Pandeeswari and Kalaiarasu, 2012). The pH of soil samples was ranged from 7.13 to 8.54 which

indicate that the soils are alkaline. Under alkaline conditions, the solubility of minerals decreases to the point that nutrient deficiencies occur. Soil EC is an easily measured yet reliable indicator of soil quality, crop performance, nutrient cycling, and biological activity and can serve as a quick indicator of plant-available nitrate-N (Johnson, 2005). EC of all the soil samples were ranged from 702 to 1247 μS/cm. Soil with EC below 400 µS/cm are considered marginally or nonsaline, while soils above 800 µS/cm are considered severely saline. The soils under analysis were found severely saline except SL8, SR 13, SR 14 and SR 17 are considered as moderately saline. TDS value ranges from 319 to 652 mg/L, higher concentration is due to leaching of salts and also domestic sewage percolate to the soil.

The value of total alkalinity is observed from 105 to 268 mg/L and increase in TA is because of mineral dissolution. Total hardness is varying from 146 to 268 mg/L and is due to the presence of calcium and magnesium. Calcium and magnesium content in the soil solution ranging between 24 to 109 mg/L and 27 to 82 mg/L respectively. Except station SL1 and SL6, Ca is in the safe range. Increase and decrease in Ca content may be their uptake by living organisms and their release on decomposition. Chloride is the most recent addition to the list of essential elements. Although chloride (Cl) is classified as a micronutrient, plants may take up as much chloride as they do secondary elements such as sulphur (Solanki, 2012). The concentration of chloride is between 163 and 310 mg/L. Fluoride shows the wide variation from 0.09 to 3.2 mg/L, above 1.0 mg/ L is not considered as safe. Sulphate and Phosphate are macronutrient elements essential for plant growth. Sulphate and phosphate ranges from 120 to 215 mg/L and 0.01 to 0.1 mg/L respectively.

Nitrogen exists in the soil system in many forms and transforms very easily from one form to another. The in and out route that N follows in the soil system is collectively called "nitrogen cycle" and is biologically influenced. The concentration of nitrate in soil samples range from 32 to 85 mg/L. Availability of Fe and Mn

content shows no marked variations and were not recorded in excess amount; these concentrations are shown in table 1. BOD is the method determines the amount of oxygen required by aerobic microorganisms to decompose the organic matter in a sample. The amount of oxygen consumed in the dilution water should range from 236 to 421 mg/L. The high values of BOD indicated the presence of an excess of biodegradable organic matter from the municipal and agricultural sewage. COD varied from 160 to 342 mg/L at the different sampling sites. The high values of COD indicate soil pollution which is related to sewage effluents discharged from town, industry or agricultural practices.

Table 2. Frequency of occurrence of microorganisms isolated from soil samples

Bacteria isolates	Frequency of occurrence (%)						
Bacillus subtilis	21						
Bacillus cereus	13						
Escherichia coli	27						
Staphylococcus aureus	21						
Pseudomonas fluorescens	4						
Klebsiella pneumoniae	10						
Staphylococcus epidermis	4						

4.2 Microbial Isolates from Soil Samples

The bacterial cultures were isolated and identified by pure culture technique, gram staining method and biochemical test methods. Bacterial cultures was isolated from thirty six soil samples, identified and maintained by sub-culturing them in nutrient broth. More than twenty five bacterial species were isolated and the dominant species are taken for account. The results of the percentage frequency of occurrence of the microbial isolates were presented in Table 2. From the results Escherichia coli (27 %) had the highest frequency of occurrence. This was followed by Bacillus subtilis and Staphylococcus aureus (21 % each) and Bacillus cereus (13 %). The lowest frequency of occurrence of 4 % was observed in Pseudomonas fluorescens and Staphylococcus epidermis respectively.

5. CONCLUSION

The present work concludes that the dyeing effluent from the industry and agricultural wastes causes the pollution problems in the surrounding environment of the sampling region. The nutrient status of the samples showed that the soil quality of the surrounding field was poor and the physicochemical characteristics of the soil also show wide variation. Soil management practices alter the physicochemical properties of soil, and the soil microbial community may respond to these changes in ways that affect the ability of the soil to resist soil-borne diseases. Soil microbial diversity drops when the soil is subjected to intensive exploitation during agricultural production. Efficient maintenance and caring of the influential soil properties are major concern in intensive crop cultivation for improving soil productive capacity, food safety and environmental quality.

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